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APPLICATION NO.	FILING DATE	FIRST NAMED INVENTOR	ATTORNEY DOCKET NO.	CONFIRMATION NO.
10/721,667	11/25/2003	Bosco P. Ho	7593-CO1	5949

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Patent Licensing Department
Nalco Company
1601 W. Diehl Road
Naperville, IL 60563-1198

EXAMINER

FERNANDEZ, SUSAN EMILY

ART UNIT	PAPER NUMBER
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1651

DATE MAILED: 09/21/2006

Please find below and/or attached an Office communication concerning this application or proceeding.

Office Action Summary

Application No.

10/721,667

Applicant(s)

HO ET AL.

Examiner

Susan E. Fernandez

Art Unit

1651

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --

Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 02 August 2006.
- 2a) ☐ This action is **FINAL**. 2b) ☒ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 1-17 is/are pending in the application.
- 4a) Of the above claim(s) _____ is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☒ Claim(s) 1-17 is/are rejected.
- 7) ☐ Claim(s) _____ is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☐ The drawing(s) filed on _____ is/are: a) ☐ accepted or b) ☐ objected to by the Examiner.
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All b) ☐ Some * c) ☐ None of:
1. ☐ Certified copies of the priority documents have been received.
 2. ☐ Certified copies of the priority documents have been received in Application No. _____.
 3. ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

* See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

- | | |
|--|---|
| 1) <input checked="" type="checkbox"/> Notice of References Cited (PTO-892) | 4) <input type="checkbox"/> Interview Summary (PTO-413)
Paper No(s)/Mail Date. _____ |
| 2) <input type="checkbox"/> Notice of Draftsperson's Patent Drawing Review (PTO-948) | 5) <input type="checkbox"/> Notice of Informal Patent Application |
| 3) <input type="checkbox"/> Information Disclosure Statement(s) (PTO/SB/08)
Paper No(s)/Mail Date _____ | 6) <input type="checkbox"/> Other: _____ |

DETAILED ACTION

A request for continued examination under 37 CFR 1.114, including the fee set forth in 37 CFR 1.17(e), was filed in this application after final rejection. Since this application is eligible for continued examination under 37 CFR 1.114, and the fee set forth in 37 CFR 1.17(e) has been timely paid, the finality of the previous Office action has been withdrawn pursuant to 37 CFR 1.114. Applicant's submission filed on August 2, 2006, has been entered.

Claims 1-17 are pending and examined on the merits.

Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

This application currently names joint inventors. In considering patentability of the claims under 35 U.S.C. 103(a), the examiner presumes that the subject matter of the various claims was commonly owned at the time any inventions covered therein were made absent any evidence to the contrary. Applicant is advised of the obligation under 37 CFR 1.56 to point out the inventor and invention dates of each claim that was not commonly owned at the time a later invention was made in order for the examiner to consider the applicability of 35 U.S.C. 103(c) and potential 35 U.S.C. 102(e), (f) or (g) prior art under 35 U.S.C. 103(a).

Claims 1-17 are rejected under 35 U.S.C. 103(a) as being obvious over Chatteraj et al. (US Pat. 6,329,165) in view of Ridgway et al. (Water Treatment Membrane Processes, McGraw Hill, 1996, pages 6.1-6.62).

Chatteraj et al. discloses a method for monitoring planktonic and sessile microbiological populations (i.e. biofouling) in an industrial water system. Chatteraj et al. teaches the same method steps as claimed in the present application, using the same fluorogenic agents (see claims 1-15; column 2, lines 30-67; column 3, lines 1-54; column 4, lines 7-34; column 5, lines 6-31, 63-64; column 6, lines 42-45; and column 7, lines 64-67 of Chatteraj et al.). Note that Chatteraj et al. indicates that commercially available fluorometers are suitable for the practice of their invention for detecting the fluorogenic agent (column 6, lines 42-45). In regards to the amount of fluorogenic agent used, Chatteraj discloses that "an effective amount of fluorogenic dye is between about 0.005 ppm and about 1.0 ppm, preferably between about 0.02 ppm and about 0.5 ppm, most preferably between about 0.04 ppm and about 0.1 ppm, and the most highly preferable amount of fluorogenic dye is 0.05 ppm" (column 5, lines 41-46). These ranges clearly fall within the ranges as claimed in claims 6-8 of the instant application.

Chatteraj et al. does not expressly disclose that: 1) the method is usable on a reverse osmosis membrane separation system such as those claimed in the instant application, and 2) biocontrol agents or biocontrol methods can be used to control biofouling (Chatteraj et al. only teach the use of oxidizing and non-oxidizing biocides, see claims 5-13).

However, Ridgway et al. teaches that membrane separation processes such as reverse osmosis play essential roles in modern water treatment practice (page 6.1, lines 1-3). Additionally, membrane biofouling is a widespread problem in these treatment processes,

Art Unit: 1651

especially in feedwater and industrial processes such as ground-water treatment, seawater desalination, and water production (i.e. in industrial water systems). See section 6.5 "Occurrence of membrane biofouling" on pages 6.20-6.21. Additionally, applicant has disclosed that membrane separation methods such as reverse osmosis are used in industrial processing of liquid streams, such as in water purification (page 1, lines 6-7 of the instant specification). Ridgway et al. also teaches that various options are available to control biofouling. Examples include the use of biocides, biocontrol agents (such as chelating agents, surfactants, and chaotropic agents), and biocontrol methods (such as ultrasound, electric fields, and air backwashes). See Table 6.4 on page 6.43.

Therefore, since reverse osmosis is commonly used in industrial water systems, as taught by Ridgway et al., it would have been obvious to use the method of Chatteraj et al. and apply it to reverse osmosis membrane separation systems. Although Chatteraj et al. only discloses the use of biocides as a means to control biofouling, it would have been obvious to modify that aspect of the method to include the use of other biocontrol agents and methods since these other venues are also known and practiced in the art, as disclosed by Ridgway et al.

Applicant's arguments filed August 2, 2006, have been fully considered but they are not persuasive. Applicant asserts that Ridgway et al. does not teach monitoring biofouling in a membrane separation system by fluorescence, as the reference teaches techniques wherein the membrane has been destroyed and extracted. However, it is respectfully pointed out that the Chatteraj reference teaches the measurement of biofouling in an industrial water system, and therefore, one of ordinary skill in the art would have been motivated to have applied the methods of Chatteraj et al. to any and all other industrial water systems, including an industrial water

Art Unit: 1651

systems comprising reverse osmosis membranes as disclosed in Ridgway et al. Ridgway et al. is provided not to teach methods of monitoring biofouling, but to teach the prevalent existence of reverse osmosis membranes in industrial water systems, and biocontrol agents and methods for controlling biofouling.

Additionally, as discussed in the previous office action, it is respectfully pointed out that it is clear that with such a modification, the fluorogenic agent would have clearly been present in the feed stream. Moreover, it is also expected that certain fluorogenic agents would have been present in the first stream and the second stream. Even if the fluorogenic agent is too large to pass through the reverse osmosis membrane, claim 1 under examination does not require that the fluorogenic agent is present in the first stream and the second stream. Specifically, instant claim 1 recites "...providing a fluorometer to detect the fluorescent signal of the fluorogenic agent in **at least one** of the feed stream, the first stream, and the second stream" (emphasis added). Note further that Chatteraj et al. discloses for the practice of its methods in-line measurement which is "one that is taken without interrupting the flow of the system being measured" (column 6, lines 64-65). Clearly, the detection of the fluorescent signal would be such that measurement is taken by the fluorometer at any point of the flow of the system, and without in interruption to the flow, and thus would have been at any of the streams. One of ordinary skill in the art would have expected that the membrane system would have fouled, and the effect of this fouling on fluorescence clearly is necessary for the invention rendered obvious by Chatteraj et al. and Ridgway et al. to accomplish its purpose. The rejections of record must be maintained.

Claims 1-14, 16, and 17 are rejected under 35 U.S.C. 103(a) as being obvious over Zeiher et al. (US 6,838,002) in view of Chatteraj et al.

Zeiher et al. discloses a method for monitoring a reverse osmosis membrane separation process wherein an inert fluorescent tracer and a tagged fluorescent agent are produced and introduced into the feed stream and a fluorometer is used to determine the amounts of the inert fluorescent tracer and the tagged fluorescent agent in at least one of the feed stream, the first stream, and the second stream (claim 1). Furthermore, at least one process parameter is evaluated based on the amounts of the inert fluorescent tracer and the tagged fluorescent agent measured (claim 2), and the amounts of these agents (claims 13-15) meet the limitations recited in instant claims 6-8. Note that Zeiher et al. discloses that the disclosed invention can be used for industrial water uses such as the use of membranes to exclude harmful microorganism from drinking water (column 25, line 10-18). Furthermore, biofouling agents may be used to treat the membranes of the system (column 20, lines 40-57).

Zeiher et al. differs from the claims under examination in that Zeiher et al. does not expressly disclose that the reverse osmosis membrane separation process being monitored is biofouling, or that the tagged fluorescent agent reacts with at least one microorganism (planktonic or sessile) within the reverse osmosis so that reacted tagged fluorescent agent is measured. Moreover, the reference does not teach that the fluorescent agent is any of those recited in instant claims 3-5, nor does it teach the determination of ratios, rate of change of a ratio, or optimal amount of biocontrol treatment. Finally, Zeiher et al. does not expressly disclose the biocontrol treatments recited in instant claims 12-14.

Chattoraj et al. discloses a method for monitoring planktonic and sessile microbiological populations (i.e. biofouling) in an industrial water system. Chattoraj et al. teaches the same method steps as claimed in the present application, using the same fluorogenic agents, and requiring the measurement of reacted fluorogenic agents and the determination of ratios, rate of change of a ratio, and optimal amounts of biocontrol treatment as recited in the claims under examination (see claims 1-15; column 2, lines 30-67; column 3, lines 1-54; column 4, lines 7-34; column 5, lines 6-31, 63-64; column 6, lines 42-45; and column 7, lines 64-67 of Chattoraj et al.). Note that Chattoraj et al. indicates that commercially available fluorometers are suitable for the practice of their invention for detecting the fluorogenic agent (column 6, lines 42-45). Chattoraj et al. also teaches the limitations relating to biocontrol recited in instant claims 11-14 (see claims 5-13 and column 7, line 60 through column 8, line 65).

At the time the invention was made, it would have been obvious to a person of ordinary skill in the art to have adapted the methods of Zeiher et al. for the assessment of biofouling as disclosed in Chattoraj et al. One of ordinary skill in the art would have been motivated to do this since Zeiher et al. teaches a known industrial water system wherein biofouling occurs. Given that Chattoraj et al. is suitable for monitoring planktonic and sessile microbiological populations in a water system, there would have been a reasonable expectation of success in using the same fluorogenic agents, steps of assessing reacted fluorogenic agents and determining optimal amount of biocontrol treatment as disclosed in Chattoraj et al. in practicing the Zeiher methods. A holding of obviousness is clearly required.

Double Patenting

The nonstatutory double patenting rejection is based on a judicially created doctrine grounded in public policy (a policy reflected in the statute) so as to prevent the unjustified or improper timewise extension of the "right to exclude" granted by a patent and to prevent possible harassment by multiple assignees. See *In re Goodman*, 11 F.3d 1046, 29 USPQ2d 2010 (Fed. Cir. 1993); *In re Longi*, 759 F.2d 887, 225 USPQ 645 (Fed. Cir. 1985); *In re Van Ornum*, 686 F.2d 937, 214 USPQ 761 (CCPA 1982); *In re Vogel*, 422 F.2d 438, 164 USPQ 619 (CCPA 1970); and, *In re Thorington*, 418 F.2d 528, 163 USPQ 644 (CCPA 1969).

A timely filed terminal disclaimer in compliance with 37 CFR 1.321(c) may be used to overcome an actual or provisional rejection based on a nonstatutory double patenting ground provided the conflicting application or patent is shown to be commonly owned with this application. See 37 CFR 1.130(b).

Effective January 1, 1994, a registered attorney or agent of record may sign a terminal disclaimer. A terminal disclaimer signed by the assignee must fully comply with 37 CFR 3.73(b).

Claims 1-5 and 9-17 are rejected under the judicially created doctrine of obviousness-type double patenting as being unpatentable over claims 1-15 of U.S. Patent No. 6,329,165 B1 (Chattoraj et al., document AG of the present IDS) in view of Ridgway et al., 1996 (document AP of the present IDS). Although conflicting claims are not identical, they are not patentably distinct from each other. Claims 1-5 and 9-17 of the instant application are directed to a method of monitoring biofouling in a membrane separation system comprising the steps of: a) adding a

Art Unit: 1651

fluorogenic agent into the system; b) allowing the fluorogenic agent to react with microorganisms (e.g. planktonic microorganisms and sessile microorganisms) in the system to form a reacted fluorogenic agent; c) providing a fluorometer to measure fluorescence; d) measuring the fluorescent signals of the fluorogenic agent and the reacted fluorogenic agent; e) monitoring biofouling in the system by determining a ratio of the fluorescent signal of the reacted fluorogenic agent to the unreacted fluorogenic agent; and f) determining the rate of change of the fluorescence ratio. The method of the instant application is limited to membrane systems such as cross-flow or dead-end flow membrane systems, examples include reverse osmosis, nanofiltration, ultrafiltration, microfiltration, inter alia. Fluorogenic agents usable in the claimed method include resazurin, 4-methylumbelliferyl phosphate, pyranine phosphate, and others. The method also comprises steps for controlling biofouling by way of biocontrol treatments such as the use of biocides, biocontrol agents or biocontrol methods. Additionally, an inert fluorescent tracer can be included.

Chattoraj et al. (in claims 1-15) discloses a method for monitoring planktonic and sessile microbiological populations (i.e. biofouling) in an industrial water system comprising the steps of: a) adding a fluorogenic agent into the system; b) allowing the fluorogenic agent to react with any planktonic microorganisms and sessile microorganisms in the system to form a reacted fluorogenic agent; c) providing a means for measuring fluorescence (i.e. a fluorometer); d) measuring the fluorescent signals of the fluorogenic agent and the reacted fluorogenic agent and calculating the ratio of the signals; e) monitoring biofouling in the system by determining a ratio of the fluorescent signal of the reacted fluorogenic agent to the unreacted fluorogenic agent; f) determining the rate of change of the fluorescence ratio; g) determining the optimal amount of

biocide to add to the system based on the ratio or rate of ratio change; and h) delivering the optimal amount of biocide into the system. Fluorogenic dyes usable include resazurin, 4-methylumbelliferyl phosphate, and pyranine phosphate. Additionally, an inert fluorescent tracer can be included. Chatteraj et al. does not expressly disclose that: 1) the method is usable on membrane separation systems such as those claimed in the instant application, and 2) biocontrol agents or biocontrol methods can be used to control biofouling (Chatteraj et al. only teach the use of oxidizing and non-oxidizing biocides). However, Ridgway et al. teaches that membrane separation processes such as reverse osmosis, microfiltration, nanofiltration and ultrafiltration, play essential roles in modern water treatment practice (page 6.1, lines 1-3). Additionally, membrane biofouling is a widespread problem in these treatment processes, especially in feedwater and **industrial** processes such as **ground-water treatment**, seawater desalination, and **water production** (i.e. in industrial water systems). See section 6.5 "Occurrence of membrane biofouling" on pages 6.20-6.21. Ridgway et al. also teaches that various options are available to control biofouling. Examples include the use of biocides, biocontrol agents (such as chelating agents, surfactants, and chaotropic agents), and biocontrol methods (such as ultrasound, electric fields, and air backwashes). See Table 6.4 on page 6.43.

Therefore, since membrane separation systems are commonly used in industrial water systems, as taught by Ridgway et al., it would have been obvious to use the method of Chatteraj et al. and apply it to membrane separation systems. Although Chatteraj et al. only disclose the use of biocides as a means to control biofouling, it would have been obvious to modify that aspect of the method to include the use of other biocontrol agents and methods since these other venues are also known and practiced in the art, as disclosed by Ridgway et al., *supra*.

Applicant's arguments filed August 2, 2006, have been fully considered but they are not persuasive. Applicant asserts that the cited references do not teach or suggest the use of fluorometers to detect fluorescence in the feed stream, first stream, and second stream. As discussed above, it would have been obvious to have modified the Chatteraj invention by including a reverse osmosis membrane separation system in the water system to be treated by practicing the Chatteraj invention as recited in claim 1 of the reference. Therefore, it is respectfully pointed out that it is clear that with such a modification, the fluorogenic agent would have clearly been present in the feed stream. Moreover, it is also expected that certain fluorogenic agents would have been present in the first stream and the second stream. Even if the fluorogenic agent is too large to pass through the reverse osmosis membrane, claim 1 under examination does not require that the fluorogenic agent is present in the first stream and the second stream. Specifically, instant claim 1 recites "...providing a fluorometer to detect the fluorescent signal of the fluorogenic agent in **at least one** of the feed stream, the first stream, and the second stream" (emphasis added). Furthermore, Chatteraj et al. also indicates that commercially available fluorometers are suitable for the practice of their invention for detecting the fluorogenic agent (column 6, lines 42-45). Note further that Chatteraj et al. discloses for the practice of its methods in-line measurement which is "one that is taken without interrupting the flow of the system being measured" (column 6, lines 64-65). Clearly, the detection of the fluorescent signal would be such that measurement is taken by the fluorometer at any point of the flow of the system, and without in interruption to the flow, and thus would have been at any of the streams. One of ordinary skill in the art would have expected that the membrane system

Art Unit: 1651

would have fouled, and the effect of this fouling on fluorescence clearly is necessary for the invention rendered obvious by Chatteraj et al. and Ridgway et al. to accomplish its purpose.

Thus, the rejections must be maintained.

Claims 1-17 are rejected under the judicially created doctrine of obviousness-type double patenting as being unpatentable over claims 1-19 of U.S. Patent No. 6,699,684. Although the conflicting claims are not identical, they are not patentably distinct from each other because the instant application reads on a membrane separation method which is recited in '684 claims 3 and 4. Therefore, the claims under examination are properly considered obvious over the patented claims. Note that no terminal disclaimer had been filed, thus the rejections must be maintained.

Claims 1-17 are provisionally rejected under the judicially created doctrine of obviousness-type double patenting as being unpatentable over claims 1-13, 16, 18-20 of copending Application No. 10/740336. Although the conflicting claims are not identical, they are not patentably distinct from each other because the instant application reads on a membrane separation method which is recited in '336 claim 3. Furthermore, while '336 includes steps for the selection of a fluorogenic agent and an inert tracer, these selection steps would have been considered a matter of routine experimentation, as it would have been obvious to determine appropriate fluorogenic agents and tracers in order to optimize the monitoring of biofouling.

This is a provisional obviousness-type double patenting rejection because the conflicting claims have not in fact been patented.

Art Unit: 1651

Therefore, the claims under examination are properly considered obvious over the '336 claims.

Note that no terminal disclaimer had been filed to obviate the rejections. Thus, the rejections must be maintained.

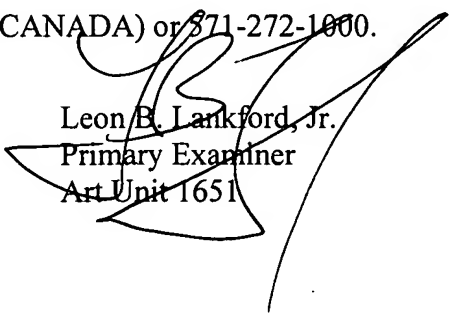
No claims are allowed.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Susan E. Fernandez whose telephone number is (571) 272-3444. The examiner can normally be reached on Mon-Fri 8:30 am - 5:00 pm.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Mike Wityshyn can be reached on (571) 272-0926. The fax phone number for the organization where this application or proceeding is assigned is 571-273-8300.

Information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see <http://pair-direct.uspto.gov>. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free). If you would like assistance from a USPTO Customer Service Representative or access to the automated information system, call 800-786-9199 (IN USA OR CANADA) or 571-272-1000.

Susan E. Fernandez
Assistant Examiner
Art Unit 1651


Leon B. Lankford, Jr.
Primary Examiner
Art Unit 1651

sef